





# New methods to detect the threat of *Phytophthora* in irrigation water



The James Hutton Institute

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#### 142 species of Phytophthora











Dave Rizzo <u>youtube.com</u> California Oak Mortality Task Force Air- or soil-borne but absolute requirement for water in all species & that provides a means of detecting **zoospores** 



#### Challenges



- Symptoms can be non-specific
- Difficult to isolate
- Species difficult to distinguish morphologically



Rhododendron with wilting symptoms caused by Phytophthora root rot.





#### **Diagnosis versus Detection**



If you already have a problem

- Conventional molecular diagnostic method
  - ➢plant material
  - DNA extraction
  - Specific PCR tests (e.g. *P. rubi, P. idaei*)
- If you want to avoid a future problem
- Detecting unknown threat
  - Irrigation water testing
  - Planting material testing
  - Keeping out new threats Biosecurity

#### Water testing can help

#### Phytophthora zoospore detection



#### Sampling water

- Irrigation water
- Water flooded through roots of pot-grown plants (factors such as timing, temperature are important)
- Rivers

#### Filtration

- Cellulose acetate filters
- DNA extraction
- PCR with *Phytophthora* species or genus specific primers
- Sequencing (high or low throughput)









## Practical application of filtration method

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- Proven detection of *P. rubi* and *P. idaei* on water flowing through coir in raspberry production systems
- Current funding to test this on large batches of plants in plant health testing stations to support plant health legislation
- Using high throughput technologies for sequencing hundreds of samples to a great depth (thousands of reads) in a single run



### **Environmental sampling**



- Four sample sites (3 = ECN)
  - Invergowrie Burn (IGB)
  - Glensaugh Burn (GSB)
  - Sourhope Burn (SRB)
  - Allt a'Mharcaidh Burn (ECN)
- Sites apparently healthy
- 10 litre water samples collected and filtered every two weeks from Dec 2011 to Feb 2014
- Filters kept cool on day of sampling and then frozen
- IGB most heavily sampled and baiting applied here only







#### Metabarcode method

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#### Journal of Microbiological Methods

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A molecular method to assess *Phytophthora* diversity in environmental samples

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• Two rounds of semi-nested PCR required

#### Illumina MiSeq



- 96 samples tagged, pooled and sequenced in a nano-cell (1 million reads)
- De-randomised error corrected resulting in 890K reads (median 8172 per sample)
- Bioinformatics pipeline...







#### **Control reaction results**

- The **PCR mix** worked well
- Inverse relation to product size
- P. fallax fewer reads



## Taxonomic summary MiSeq





- Majority of reads *Phytophthora*
- Almost 30% downy mildew species
- 8 % novel mostly new Phytophthoras
- Tiny proportion of other reads so primers highly specific

# Downy mildew summary MiSeq





Hyaloperonospora Parasitica (Brassica)  37 species - a lot of diversity

- 27 Peronospora
- 2 Hyaloperonospora
- 1 Bremia (lactucae)
- 3 Pseudoperonospora
- 4 Plasmopora
- Range of obligate species seems logical for plant hosts likely in these environments







- 45 known species
- *P. infestans*? 85% of these in GSB and generally weaker samples
- *P. gonapodyides* most abundant phylotype

## Illumina results (overview)





- Clear site-to-site variation
- Less diversity at GSB Glensaugh (mostly P. infestans)
- Most diversity at Invergowrie (more samples and lowland)

# Illumina results (growing season)



- Growing season (April Sept) No\_Grow (low\_grow) Oct Mar
- P. gonapodyides dominates at each. P. cambivora in both
- More *P. syringae* over winter
- More Pseudoperonospora urticae and Phy. Sp. over summer





- *P. gonapodyides* dominates at each time point
- Regular occurrence of some species
- Periodic or sporadic appearance of others





The method works well but...

- Laboratory time still takes weeks to run and analyse the data
- Bioinformatics pipeline needs work to increase speed and confidence
- Costs? Approx. £2000 a run of 96 samples but the costs will fall