

Potato Council-Funded Blackleg Research Proposal (*Pectobacterium* spp.)

Contamination of high grade potato seed stocks by *Pectobacterium* species and the effects of sulphuric acid treatment on pathogen spread

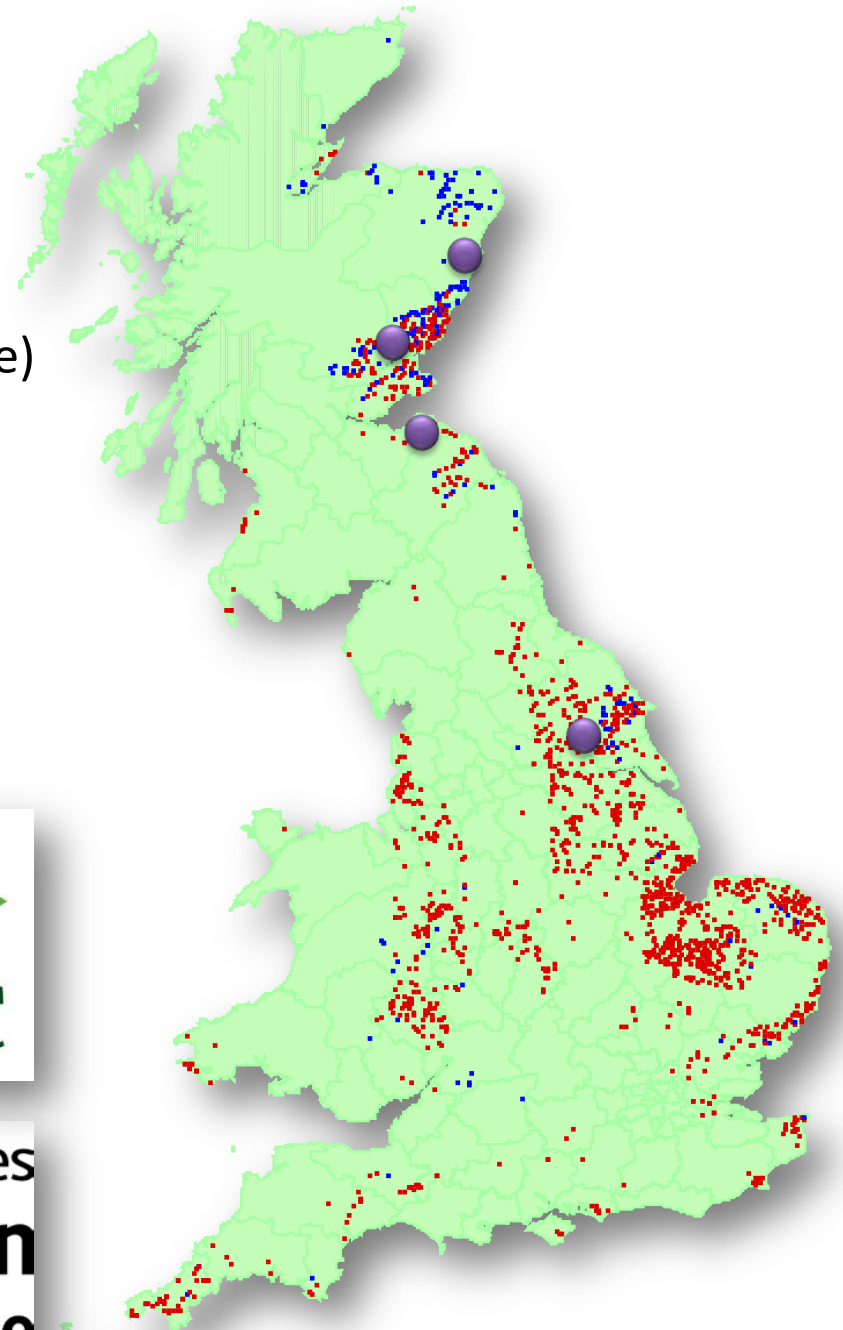


The James
Hutton
Institute



Participants

- Ian Toth **James Hutton Institute** (Dundee)
- Gerry Saddler **SASA** (Edinburgh)
- Stuart Wale **SRUC** (Aberdeen)
- John Elphinstone **Fera** (York)

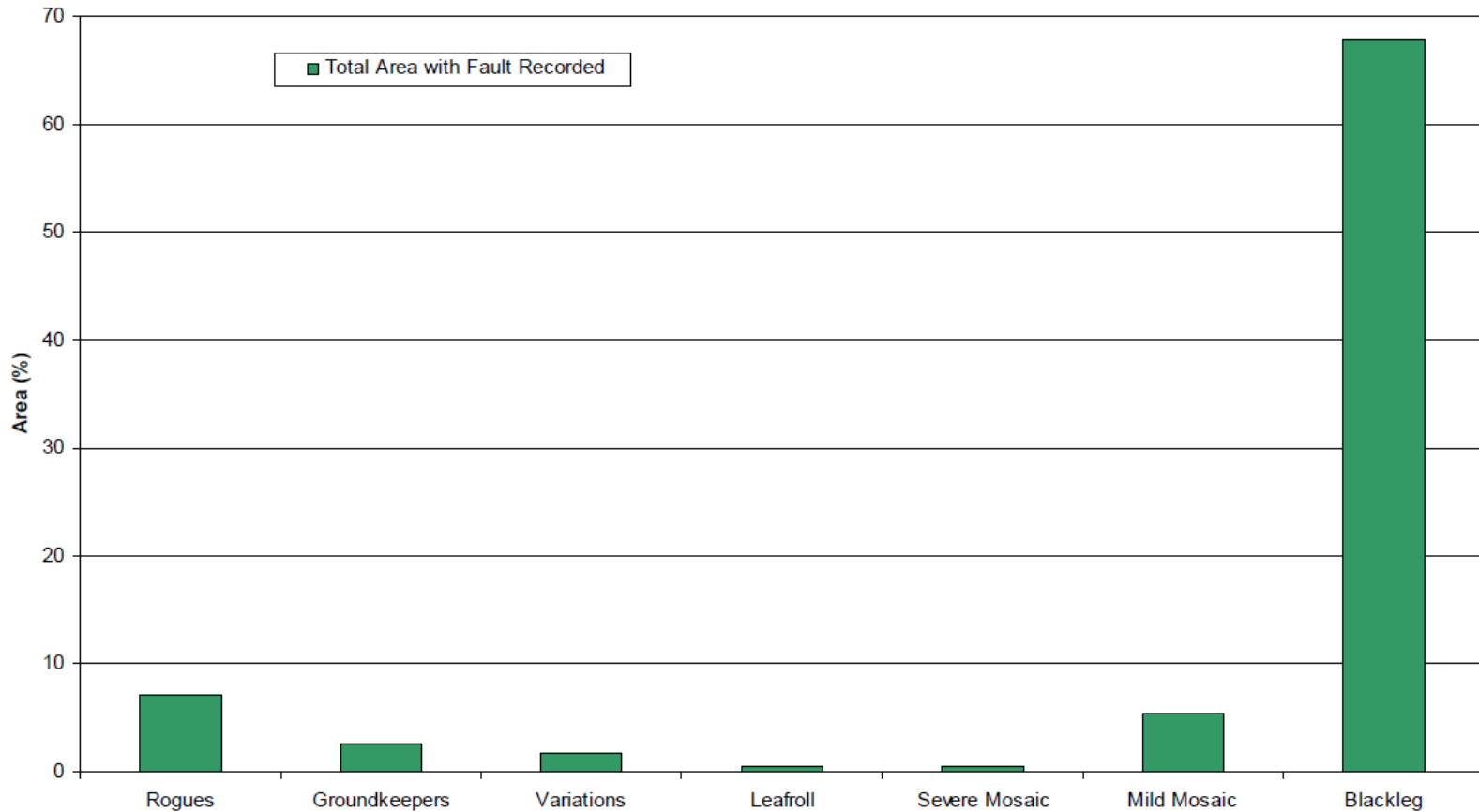


Blackleg and seed production in Scotland



Fig. 2 Total area with fault recorded at Growing Crop Inspection 2012

Figures expressed as % of seed area entered for inspection

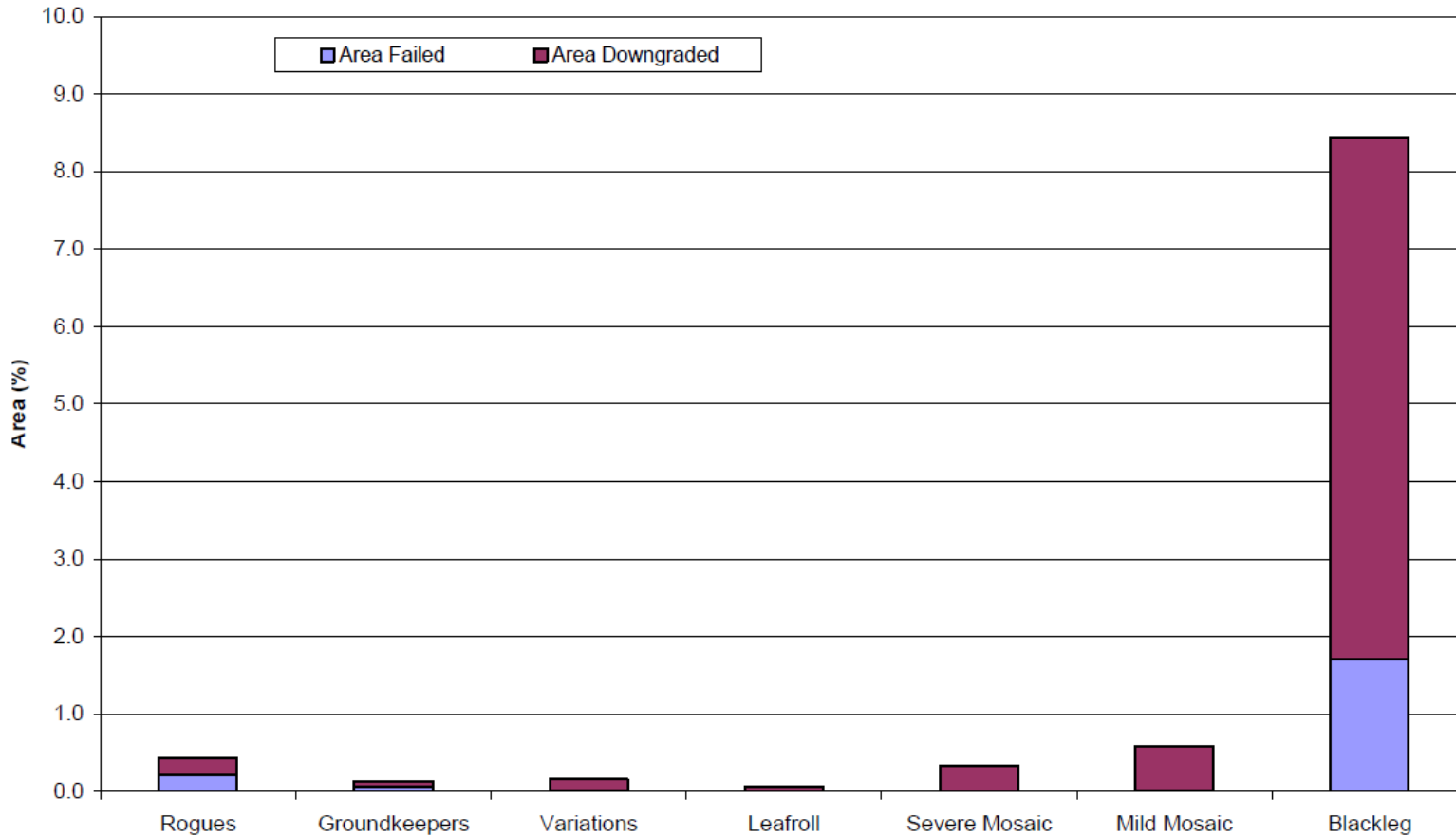


SASA © Crown Copyright

Blackleg and seed production in Scotland



Fig. 4 Seed area downgraded and failed at Growing Crop Inspection 2012 by fault
Figures expressed as % of seed area entered for inspection

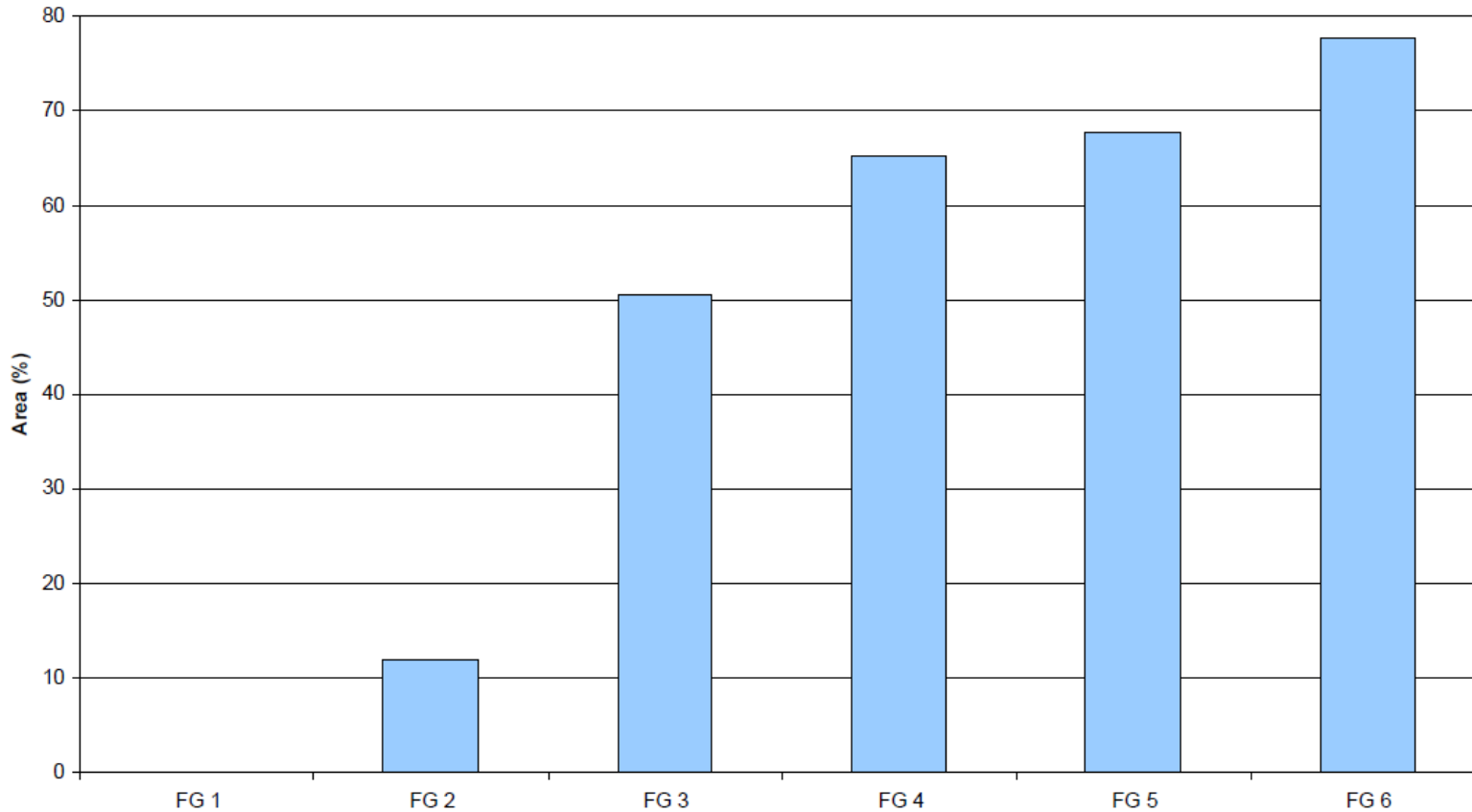


SASA © Crown Copyright

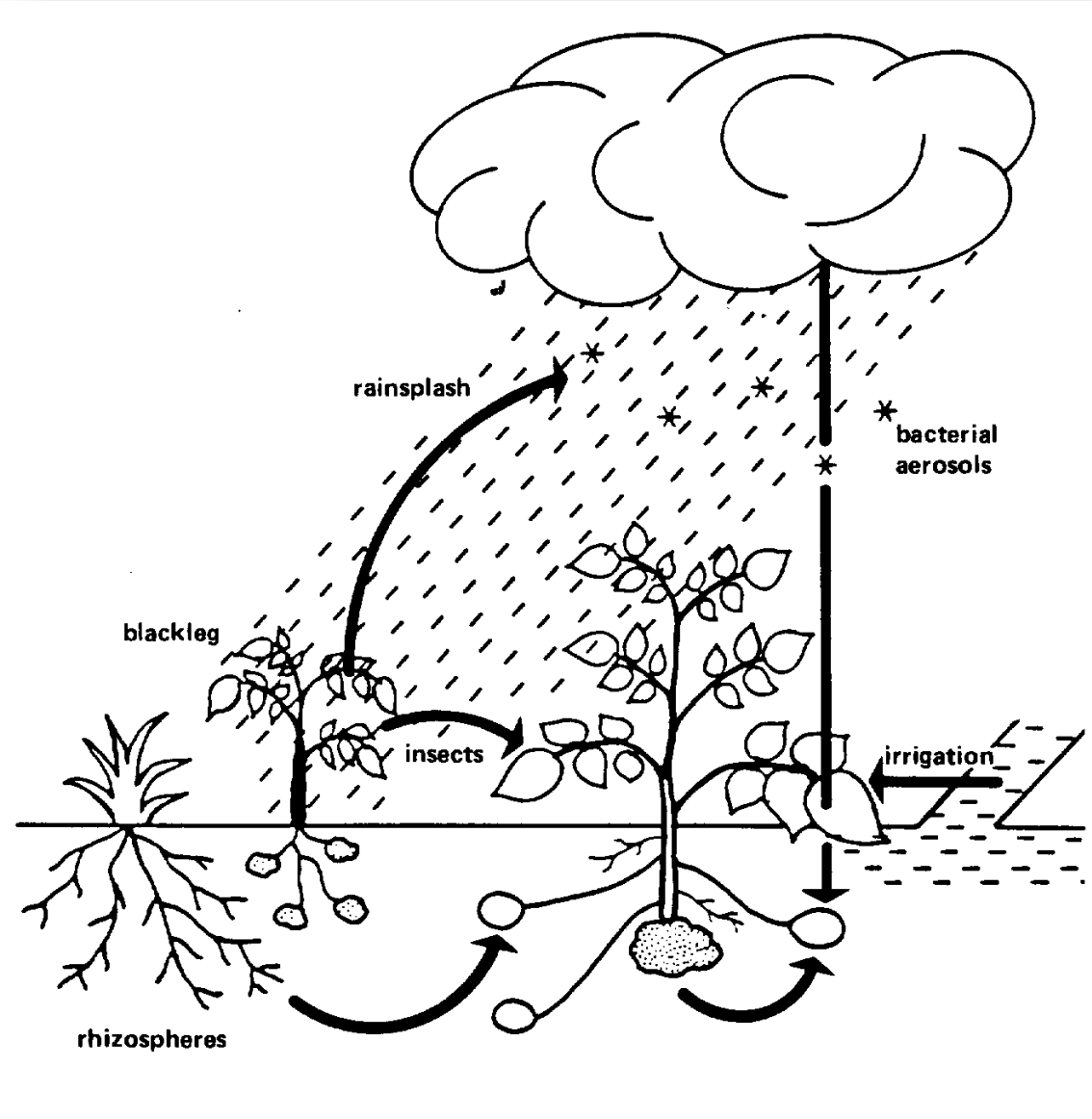
Blackleg and seed production in Scotland



Fig. 6 Area with blackleg recorded for each field generation (FG)
Figures expressed as % of seed area entered for inspection for each field generation



Contamination of pre-basic seed (PB1)



Aims

- How /when do **early field generations** become contaminated by *Pectobacterium* spp. (particularly *P. atrosepticum* - *Pba*)
- Use this information to modify current control strategies
- Delay infection in subsequent field generations
- Compare currently available haulm desiccants with sulphuric acid in their effect on spread of *Pba*.



Contamination of PB1 crops

- Monitor pre-basic crops from planting as mini-tubers to identify when *Pectobacterium* contaminates haulm, roots / tubers.

Key stages of PB1 crops monitored:

- Mini-tubers pre-planting
- 3, 6, 9 and 12 weeks after emergence
- Just prior to haulm destruction
- 1 week after last haulm treatment
- Harvest
- Storage

























Contamination of PB1 crops

- In two seasons very little contamination of haulm was detected but *Pectobacterium* was found on below ground parts
- Levels of contamination, above and below ground, was strongly influenced by environmental conditions
- There was a strong indication that blackleg in a PB field can affect the level of contamination of PB1 in the same field



VNTR profiles of *Pba* isolates

Location and seed stock	 Harvester before lifting	 Blackleg plants	 Harvester after lifting	 Harvested tubers
Scotland Farm 2 Variety 1 PB-2				
Scotland Farm 2 Variety 2 PB-2				
Scotland Farm 2 Variety 3 PB-2				
Scotland Farm 3 Variety 4 PB-2				
Scotland Farm 4 Variety 5 PB-2				
Scotland Farm 5 Variety 6 PB-2				

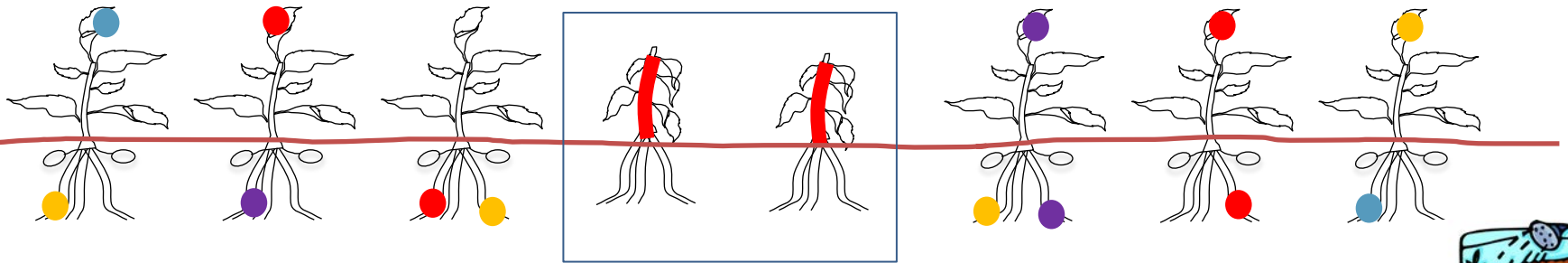
VNTR profiles: 1  2  3  4  5  6  7  8  9  10  11  12  13  14 

Sources of *Pba* contamination in PB1 crops

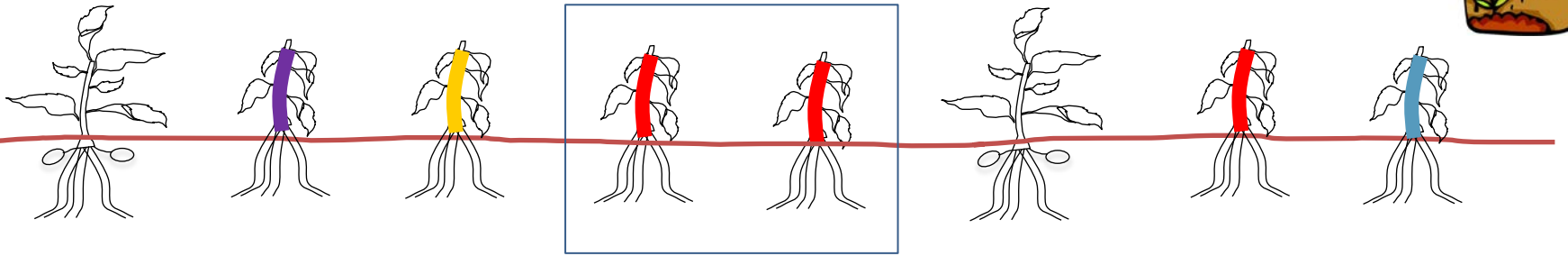
- DNA fingerprinting methods have been developed to fingerprint strains of *Pba*.
- Grading /harvesting equipment picks up *Pba* from infected crops but this *Pba* is not the only source of *Pba* contamination detected on harvested tubers.
- *Pba* strain types isolated from blackleg plants in neighbouring seed crops contribute to contamination and even blackleg disease in PB1 and PB2 crops.



9



12

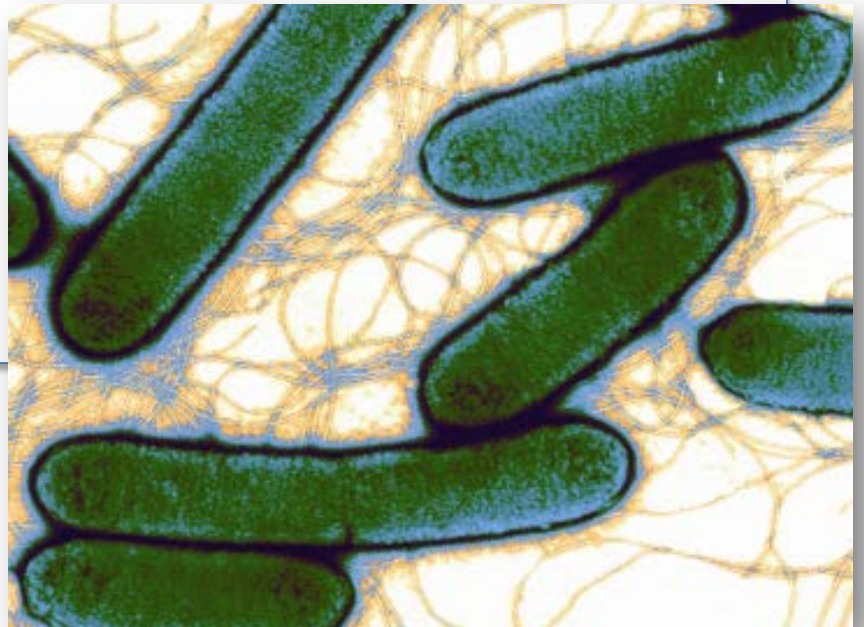


Experimental field plots

- *Pba* can spread from diseased plants in a field to other plants in that field.
- Isolates on plants are found both above and below ground.
- Highest numbers of *Pba* appear on roots and stems, where they may multiply and infect.
- Even with a heavy inoculum from artificially infected blackleg plants, natural *Pba* and other *Pectobacterium* species contaminate plants at increasingly high levels and can cause disease.
- Harvested tubers have both surface (including lenticels) (2/3) and systemic (1/3) contamination.

Determining populations

- Determine whether the population of *Pba* has changed/is changing over time
 - Relate particular populations to sources of inoculum
 - Determine whether **other *Pectobacterium* spp.** cause of blackleg in GB
 - *P. carotovorum*
 - *P. brasiliense*
 - *P. wasabiae*



Blackleg findings in seed potato stocks entered for classification in England and Wales

	2010	2011	2012	2013	2014
% seed stocks with blackleg	32.1	21.5	33.8	29.5	28.6
% blackleg caused by <i>D. solani</i>	7.0	2.3	1.8	1.7	0.4
% blackleg caused by <i>D. dianthicola</i>	0.4	0.6	1.8	0.0	0.4
% blackleg caused by <i>P. atrosepticum</i>	75.2	74.4	84.1	86.5	81.2
% blackleg caused by other <i>Pectobacterium</i> spp.	17.4	22.7	12.3	11.8	18.0

Causes of blackleg in England / Wales

- 20% of blackleg in seed crops in England and Wales caused by
 - *P. carotovorum* subsp. *carotovorum* (*Pcc*)
(mixed origins incl. GB)
 - *P. carotovorum* subsp. *brasiliense* (*Pcb*)
(all NL origin)
 - *P. wasabiae* (*Pwa*)
(mixed origins incl. GB)
- *Dickeya solani* and *Dickeya dianthicola* are minor causes of blackleg (<1% of seed stocks in 2014).
- Whereas *Pba*, *Pcc*, and *Pwa* are already widely distributed in European seed, *Pcb* appears to be recently emerging.



Blackleg findings in seed potato stocks entered for classification in England and Wales

Year	Number of crops studied	Number of positive findings	
		<i>Dickeya</i>	<i>Pwa</i>
2007	22	0	0
2008	2	0	0
2009	174	4	0
2010	545	9	6
2011	752	0	8
	+7 groundkeepers from 2009 + 2010	0	
2012	821	0	8
	+11 groundkeepers from 2009 + 2010	0	
2013	671	0	0
	+6 groundkeepers from 2009 + 2010	0	
2014	664	0	1

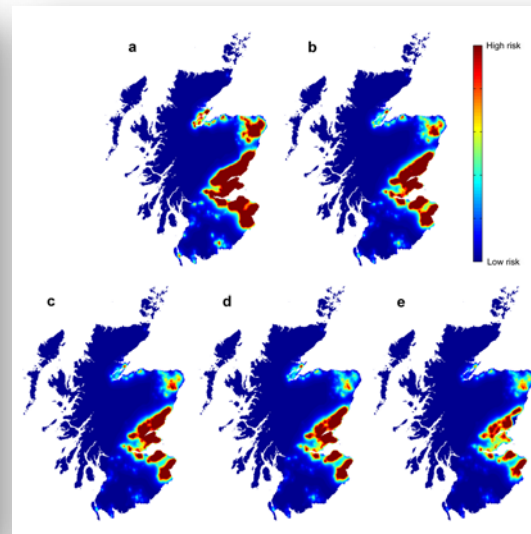
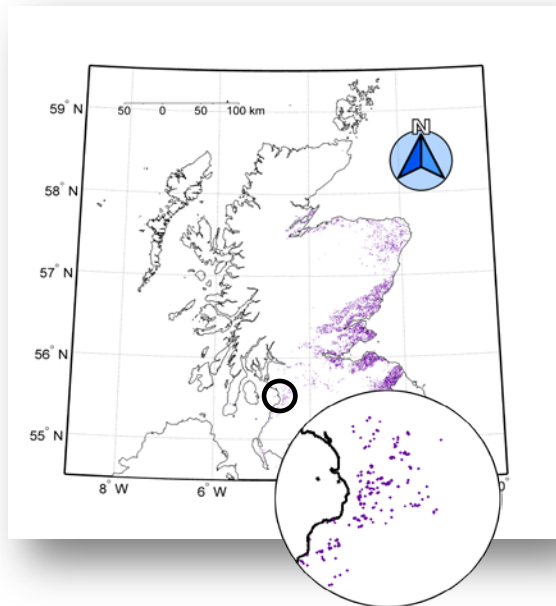
Pectobacterium species in Scotland

- In Scotland *P. wasabiae* is present in small numbers and *P. carotovorum* subsp *brasiliense* has been found in stocks imported into Scotland in 2006-2008.



Modelling the spread

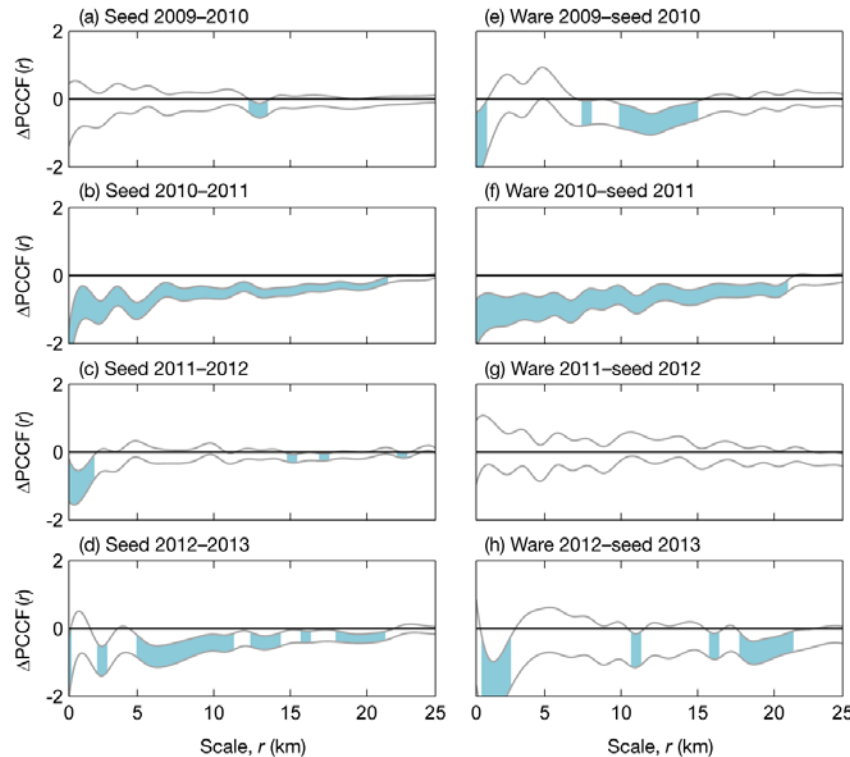
- Does planting high grade seed close to blackleg infected crops increase the risk of contamination?
- Using SPUDS database and across landscapes modelling
- Assess progeny crops historically, and going forward, to identify whether this increases the risk of blackleg in subsequent generations.



Spread risk model

Changes over space and time

- Incidences of blackleg appear to cluster and show some degree of clustering with the location of ware crops and with seed crops exhibiting blackleg disease both within an year and between years.



Effectiveness of sulphuric acid

- To compare effectiveness of sulphuric acid in comparison with current standard haulm desiccation programmes to determine their relative impact on *Pba* spread and infection of progeny tubers
 - Small plot trial using cv. Desiree mini-tubers
 - Plants inoculated with marked *Pba* strain 1 week before haulm destruction.

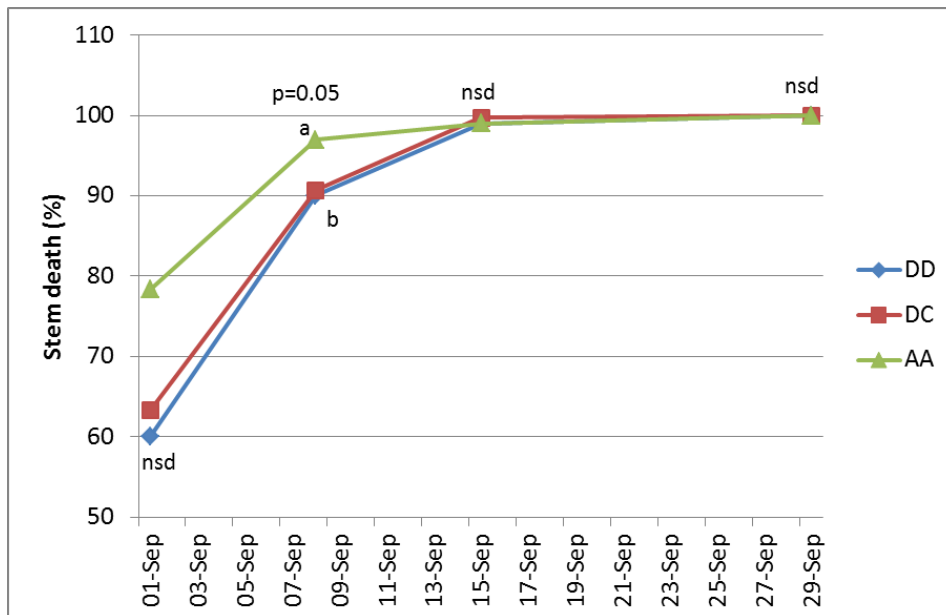


Effectiveness of sulphuric acid

- Three haulm destruction treatments applied:
 - Diquat
 - Diquat followed by Carfentrazone
 - Sulphuric acid (a 77% solution of clean 'sulphur burnt').
- Measure *Pba* contamination before and after harvest, on foliage, stems, stolons, roots and tubers (lenticel and vascular)
- Monitor effectiveness of the haulm destruction treatments, - degree of stem desiccation and speed of haulm and pathogen kill

Effectiveness of sulphuric acid

- Sulphuric acid provided more rapid haulm kill than current desiccant options (Diquat / Diquat-Carfentrazone)
- Evidence that sulphuric acid can reduce levels of contamination of *Pba* on potato plants



Recommendations

- Monitoring of seed health should take into account the diversity of the blackleg bacteria.
- Separation of PB fields for years 1 & 2.
- Isolation of PB fields may reduce contamination from outside sources (space / polytunnels?).
- When undertaking diagnostics, test for contamination on both the tuber surface (lenticels) and systemically.
- Keep irrigation of all seed crops to a minimum.
- Harvest high grade stocks as early as possible.

Acknowledgements

